

A Mild and Convenient Procedure for the Conversion of Toxic β -Asarone into Rare Phenylpropanoids: 2,4,5-Trimethoxycinnamaldehyde and γ -Asarone[†]

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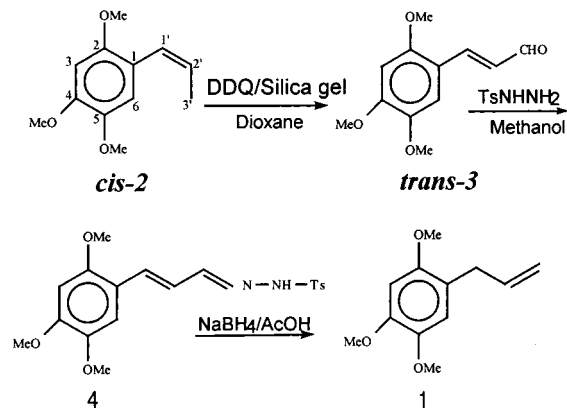
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Oxidation of β -asarone (**2**) with DDQ gave *trans*-2,4,5-trimethoxycinnamaldehyde (**3**), which on treatment with *p*-toluenesulfonyl hydrazine provided corresponding α,β -unsaturated hydrazone derivative (**4**). Reduction of **4** with sodium borohydride in acetic acid afforded γ -asarone (**1**) in 43% yield.

In nature asarones¹ exist in three isomeric forms, namely, α -, β -, and γ -asarone (*trans*-2,4,5-trimethoxy-1-propenylbenzene, *cis*-2,4,5-trimethoxy-1-propenylbenzene, and 1-allyl-2,4,5-trimethoxybenzene, respectively). γ -Asarone (**1**) is a rare phenylpropanoid² first isolated from *Caesulia axillaries*³ and later detected as a biologically active⁴ constituent of various essential oil bearing plants.⁵ However, no simple method is available for separation and isolation of asarones (α , β , and γ) in single isomeric form due to their similar physical properties. Separation of less abundant **1** via column chromatography of asarones-rich essential oil is particularly difficult. A few methods for the synthesis of **1** are found in the literature,^{2,6} but protocols either involve multiple steps starting from dimethoxyphenol^{6a,c} with overall poor yield or require careful handling of sodium perchlorate during electrolysis of methyl eugenol.^{2,6b} Herein we report a simple synthesis of **1** from toxic β -asarone⁷ (**2**) via formation of a key intermediate, 2,4,5-trimethoxycinnamaldehyde (**3**), as outlined in Scheme 1; compound **3** is itself a rare phenylpropanoid found in traces in *C. axillaries*⁸ and *Alpinia flabella*.⁹

β -Asarone (**2**) is found in several plants¹ including *Acorus calamus*¹⁰ (family Araceae). The high percentage of toxic **2** (varying from 70 to 90% in tetraploid and hexaploid strains¹¹ distributed extensively in India, Pakistan, Bangladesh, Japan, and China) restricts the market potential¹² of calamus oil. Therefore, as a part of our continuing efforts to generate value-added products,¹³ compound **2** has been converted into rarer phenylpropanoids **3** and **1**. Treatment of **2** (*cis*-isomer) with DDQ¹⁴ in wet dioxane afforded **3** in 41% yield, whereas the addition of a catalytic amount of silica gel (60–120 mesh size) improved the yield of **3** up to 62%. ¹H NMR clearly indicated the formation of **3** as *trans* α,β -unsaturated aldehyde in which the olefinic proton appeared at δ 7.81 ($J = 15.8$ Hz) rather than the expected *cis*-isomer.¹⁵ Alternatively, the above oxidation was repeated starting with *trans*-asarone, which produced an 84% yield of expected *trans*-cinnamaldehyde (**3**), identified by mixed melting point and comparison of its spectral data with an authentic sample.^{8,13c} These results clearly indicated formation of the thermodynamically more stable *trans* isomeric form of cinnamaldehyde (**3**) whether starting from *cis*- or *trans*-asarone; however, *trans*-asarone provided higher yields. Compound **3** has been isolated, so far, from two plants,^{8,9} but only in trace amounts. Treatment of a methanolic solution of **3** with *p*-toluenesulfonylhydrazine¹⁶ (1.2 equiv) afforded the corresponding α,β -unsaturated

Scheme 1



tosylhydrazone derivative (**4**) in 79% yield. To obtain the product **1**, the reduction and double-bond migration¹⁷ of tosylhydrazone derivative **4** proceeded smoothly and in good yield (43%) with acetic acid–sodium borohydride, but yields were lower using sodium cyanoborohydride–sulfolane. The R_f value (0.39 in 4% ethyl acetate in hexane) of **1** was exactly the same as that for **2**; however, spectral data of **1** were found consistent with those in the literature.^{1,6c} Finally, we conclude that the synthesis of **3** and **1** from **2** is convenient and efficient in comparison to reported^{2,6,8} methods.

Experimental Section

General Experimental Procedures. Melting points were determined with a Mettler FP80 micromelting point apparatus and are uncorrected. NMR spectra were recorded on a Bruker AM-300 spectrometer in CDCl₃ using TMS as an internal standard and EIMS on a JEOL JMS-HX 300 mass spectrometer, respectively. DDQ and α -asarone were purchased from Merck and Sigma Chemical Co., respectively.

Plant Material and Isolation of β -Asarone (2**).** The rhizomes of *Acorus calamus* were collected from Palampur (1300 m altitude) in May–June 1997, and the plant specimen was compared against a voucher specimen (no.1066) in the herbarium of the IHBT, Palampur, India. The steam distillation of rhizomes gave calamus oil (1.7% w/w), which after column chromatography on a silica gel column with hexane/ethyl acetate (99:1 to 90:10) provided **1** (82% w/w) as pale yellow liquid (R_f 0.39 on silica gel TLC plate in 4% ethyl acetate in hexane), and its spectral data agreed well with reported^{1,13} literature values.

Preparation of 2,4,5-Trimethoxycinnamaldehyde (3**) from β -Asarone (**2**).** A mixture of **2** (2.08 g, 0.01 mol) and DDQ (4.54 g, 0.02 mol) in wet dioxane (40 mL) was stirred for 15 min. A catalytic amount of silica gel (0.2–0.3 g) was added

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to the above mixture with constant stirring at room temperature overnight. The precipitated hydroquinone (DDQH₂) was filtered and further washed with dioxane. The filtrate and washings were concentrated to dryness, resuspended in CHCl₃ (25 mL), washed with H₂O (2 × 10 mL), NaHCO₃ (10%, 2 × 10 mL), and brine (2 × 10 mL), and dried over anhydrous Na₂SO₄. The residue obtained on evaporation of the solvents was column chromatographed on silica gel containing some neutral alumina at the top. The column was eluted with hexanes–ethyl acetate (9:1 to 3:2). The fractions were monitored on a TLC plate, and the desired fractions were combined and solvent removed under vacuum to afford **3** (1.38 g) in 62% yield as a yellow solid with *R*_f 0.67 (25% ethyl acetate in hexane): mp 139 °C (lit.⁸ 140 °C); ¹H NMR (CDCl₃, 300 MHz) δ 9.65 (1H, d, *J* = 7.8 Hz, H-3'), 7.81 (1H, d, *J* = 15.8 Hz, H-1'), 7.03 (1H, s, H-6), 6.64 (1H, dd, *J* = 15.8 Hz, *J* = 7.8 Hz, H-2'), 6.51 (1H, s, H-3), 3.95 (s, 3H, 2-OCH₃), 3.91 (s, 3H, 4-OCH₃), 3.87 (s, 3H, 5-OCH₃); ¹³C NMR (CDCl₃, 75.4 MHz) δ 194.1 (C-3'), 154.1 (C-1'), 153.2 (C-2), 147.6 (C-4), 143.3 (C-5), 126.4 (C-2'), 114.5 (C-1), 110.5 (C-6), 96.5 (C-3), 56.4 (5-OCH₃), 56.2 (2-OCH₃), 56.0 (4-OCH₃); EIMS *m/z* 222 [M]⁺ (44), 207 (18), 191 (100), 179 (14), 171 (27), 151 (14), 147 (7), 69 (58), 58 (80).

Preparation of 2,4,5-Trimethoxycinnamyltosylhydrazide (4). The cinnamaldehyde **3** (1.11 g, 0.005 mol) was dissolved in boiling MeOH (40 mL), and the solution was cooled to room temperature. *p*-Toluenesulfonylhydrazine (1.12 g, 0.006 mol) was added and the solution stirred at room temperature overnight. The red viscous material obtained on evaporation of solvents was chromatographed on a silica gel column with hexanes–ethyl acetate (9:1 to 3:7) as the eluent. The fractions containing **4** were pooled on the basis of TLC. Evaporation of solvent gave yellow crystals (1.54 g) of tosylhydrazide (**4**) in 79% yield with *R*_f 0.32 (25% ethyl acetate in hexane): mp 155–168 °C; ¹H NMR (CDCl₃) δ 7.88 (2H, d, Ts-H), 7.62 (1H, d, H-3'), 7.32 (2H, d, Ts-H), 7.02 (1H, d, H-1'), 6.98 (1H, s, H-6), 6.69 (1H, dd, H-2'), 6.49 (1H, s, H-3), 3.96 (3H, s, 2-OCH₃), 3.93 (3H, s, 4-OCH₃), 3.86 (3H, s, 5-OCH₃), and 2.43 (3H, s, Ts-CH₃).

Preparation of γ -Asarone (1). A solution of tosylhydrazide **4** (0.78 g, 0.002 mol) in glacial acetic acid (8 mL) was added dropwise to a precooled solution of sodium borohydride (0.38 g, 0.01 mol) and acetic acid (3 mL) under nitrogen atmosphere. The reaction mixture was stirred initially at 5–10 °C for 1 h and finally at 90–120 °C for 12 h. The mixture was poured on to ice-cooled H₂O and extracted with CH₂Cl₂ (20 mL × 3). The organic layers were combined, washed with dilute sodium hydroxide and saturated brine, and dried over anhydrous Na₂SO₄. The crude product was chromatographed on silica gel (hexanes–ethyl acetate from 99:1 to 90:10) to

obtain 0.18 g (43%) of **1** as a viscous liquid with *R*_f 0.39 (4% ethyl acetate in hexane): ¹H NMR (CDCl₃) at δ 6.70 (1H, s, H-6), 6.53 (1H, s, H-3), 5.95 (1H, m, H-2'), 5.04 (2H, m, H-3'), 3.88 (3H, s, 2-OCH₃), 3.83 (3H, s, 4-OCH₃), 3.80 (3H, s, 5-OCH₃), and 3.32 (2H, d, *J* = 6.6 Hz, H-1'); ¹³C NMR (CDCl₃) δ 151.72 (C-2), 148.30 (C-4), 143.40 (C-5), 137.04 (C-2'), 120.42 (C-1), 115.58 (C-3'), 114.38 (C-6), 98.41 (C-3), 56.99 (4-OCH₃ & 5-OCH₃), 56.62 (2-OCH₃), and 34.05 (C-1'); EIMS *m/z* 208 [M]⁺ (100), 193 (60), 165 (40), 134 (10), 77 (18), 69 (34). On the basis of the above spectral data and comparing with reported literature,^{1,6c} the liquid was identified as **1**.

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References and Notes

- Patra, A.; Mitra, A. K. *J. Nat. Prod.* **1981**, *44*, 668–669.
- Vargas, R. R.; Pardini, V. L.; Viertler, H. *Tetrahedron Lett.* **1989**, *30*, 4037–4040.
- Devgan, O. N.; Bokadia, M. M. *Aust. J. Chem.* **1968**, *21*, 2001.
- Masuda, T.; Inazumi, A.; Yamada, Y.; Padolina, W. G.; Kikuzaki, H.; Nakatani, N. *Phytochemistry* **1991**, *30*, 3227–3228.
- Avella, E.; Diaz, P. P.; Diaz, A. M. P. de; De-Diaz, A. M. P. *Planta Med.* **1994**, *60*, 195.
- (a) Shulgin, A. T. *Can. J. Chem.* **1965**, *43*, 3437. (b) Shaopeng, W.; John, S. S. *Tetrahedron Lett.* **1990**, *31*, 1513–1516. (c) Marina, D. G.; Pietro, M.; Antonino, P.; Lucio, P. *Phytochemistry* **1992**, *31*, 4119–4123.
- (a) Taylor, J. M.; Jones, W. I.; Hogan, E. C.; Gross, M. A.; David, D. A.; Cook, E. L. *Toxicol. Appl. Pharmacol.* **1967**, *10*, 405. (b) Keller, K.; Odenthal, K. P.; Leng, P. E. *Planta Med.* **1985**, *1*, 6–9. (c) Abel, G. *Planta Med.* **1987**, *53*, 251–253.
- Kulkarni, M. M.; Sohoni, J.; Rojarkar, S. R.; Nagasampagi, B. A. *Ind. J. Chem. Sect. B* **1986**, *25B*, 981–982.
- Kikuzaki, H.; Tesaki, S.; Yonemori, S.; Nakatani, N. *Phytochemistry* **2001**, *56*, 109–114.
- Motley, T. J. *Econ. Bot.* **1994**, *48*, 397–412.
- Mazza, G. *J. Chromatogr.* **1985**, *328*, 179–206.
- Harborne, J. B.; Baxter, H. In *Phytochemical Dictionary, A Handbook of Bioactive Compounds from Plants*; Taylor & Francis Ltd: Washington, DC, 1993; p 474.
- (a) Sinha, A. K. US Patent No. 09-652376 filed on August 31, 2000. (b) Sinha, A. K.; Joshi, B. P.; Dogra, R. *Nat. Prod. Lett.* **2001**, *15*, 439–444. (c) Sinha, A. K.; Dogra, R.; Joshi, B. P. *Indian J. Chem.* **2002**, *41B*, in press.
- Tomrny, I.; Shimming, L.; Knut, L. *Tetrahedron Lett.* **1998**, *39*, 2413–2416.
- Saxena, D. B. *Phytochemistry* **1986**, *25*, 553–555.
- Goffredo, R.; Alessandro, M. *Synthesis* **1976**, 530–532.
- (a) Hutchins, R. O.; Kacher, M.; Rua, L. *J. Org. Chem.* **1975**, *40*, 923–926. (b) Hutchins, R. O.; Nicholas, R. N. *J. Org. Chem.* **1978**, *43*, 2299–2301.

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